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Thiazole, Oxadiazole, and Carboxamide Derivatives of Artemisinin are Highly Selective and Potent Inhibitors of *Toxoplasma gondii*

Christopher P. Hencken,*[,†] Lorraine Jones-Brando,[‡] Claudia Bordón,[‡] Remo Stohler,[†,∥] Bryan T. Mott,[†] Robert Yolken,[‡] Gary H. Posner,[†,§] and Lauren E. Woodard[†]

[†]*Department of Chemistry, The Johns Hopkins University, 3400 North Charles Street, Baltimore, Maryland 21218,* [‡]*Stanley Division of Developmental Neurovirology, Department of Pediatrics, Johns Hopkins University School of Medicine, 600 North Wolfe Street, Baltimore, Maryland 21287, and* [§]*Johns Hopkins University Malaria Research Institute, Bloomberg School of Public Health, Baltimore, Maryland 21205.* [∥] *Present Address: Syngenta Crop Protection Münchwilen AG, Breitenloh 5, 4333 Münchwilen, Switzerland*

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We have prepared 23 new dehydroartemisinin (DART) trioxane derivatives (11 thiazoles, 2 oxadiazoles, and 10 carboxamides) and have screened them for in vitro activity in the *Toxoplasma* lytic cycle. Fifteen (65%) of the derivatives were noncytotoxic to host cells ($TD_{50} \geq 320\ \mu M$). Eight thiazole derivatives and two carboxamide derivatives displayed effective inhibition of *Toxoplasma* growth ($IC_{50} = 0.25-0.42\ \mu M$), comparable in potency to artemether ($IC_{50} = 0.31\ \mu M$) and >100 times more inhibitory than the currently employed front-line drug trimethoprim ($IC_{50} = 46\ \mu M$). The thiazoles as a group were more effective than the other derivatives at inhibiting growth of extracellular as well as intracellular parasites. Unexpectedly, two thiazole trioxanes (**5** and **6**) were parasiticidal; both inhibited parasite replication irreversibly after parasite exposure to 10 μM of drug for 24 h, whereas the standard trioxane drugs artemisinin and artemether were not parasiticidal. Some of the new derivatives of artemisinin described here represent effective anti-*Toxoplasma* trioxanes as well as molecular probes for elucidating the mechanism of action of the DART class of artemisinin derivatives.

Introduction

Toxoplasma gondii is an obligate, intracellular, apicomplexan protozoan with worldwide distribution. The complete life cycle of *T. gondii* comprises two phases: sexual and asexual. These phases involve several distinct life forms: tachyzoites, bradyzoites, merozoites, and sporozoites. Tachyzoites and merozoites function largely to expand the parasite population within the host while bradyzoites and sporozoites are capable of spreading infection to new hosts via the environment.[1] The complete life cycle takes place only within members of the Felidae family, making them the definitive hosts. Only the asexual phase takes place inside intermediate hosts, any warm-blooded animal including humans. All hosts are infected by ingesting sporulated oocysts shed into the environment by infected cats by consuming bradyzoites in the form of tissue cysts from infected animals, or by drinking water contaminated with either of these.[2] Human fetuses also can become infected transplacentally from their mother.

Globally, estimates are that one person in three is infected with *T. gondii*.[3] In the United States, nearly 25% of the adult population has been infected with this organism. The percentage of people infected correlates with demographic factors such as age, race, and socioeconomic status and environmental factors such as ambient temperature, humidity, and altitude. In some areas of the world, the seropositivity in adults ranges from 80%[4,5] to 95%.[6]

T. gondii is implicated in several maladies in humans including encephalitis, spontaneous abortions in pregnant women, and ocular disease. Recently, a report was published linking *Toxoplasma* infection with schizophrenia.[7] Infected immunocompetent adults rarely experience acute symptoms beyond fever, malaise, and adenopathy. Individuals with HIV/AIDS, cancer chemotherapy patients, or those with otherwise compromised immune systems can experience neurologic, ocular, or systemic toxoplasmosis with widespread organ damage. The potential impact of the disease is sizable considering the near 50 million people with HIV/AIDS or cancer, accounting for almost one percent of the total world population.[8,9] Toxoplasma infection can lead to seizures and life-threatening illnesses such as encephalitis in immunocompromised individuals and, if left untreated, can be fatal.[10]

Current therapies for treating *Toxoplasma* infections show limited efficacy and are often associated with severe side effects. These therapies include inhibition of folate metabolism, of protein synthesis, and of electron transport. Antifolate combination therapies employing diaminopyrimidines, such as trimethoprim or pyrimethamine, combined with sulfonamides, such as sulfadiazine or sulfamethoxazole, act synergistically against various bacterial and parasitic microorganisms. Protein synthesis inhibitors, such as macrolide and lincosamide antibiotics, are a second class of medications with anti-*Toxoplasma* activity. Their mechanism of action in *T. gondii* is assumed to inhibit plastid or mitochondrial organellar protein synthesis. The third class of anti-*Toxoplasma* drugs comprises the electron-transport inhibitors such as atovaquone. Atovaquone is occasionally used with pyrimethamine

*To whom correspondence should be addressed. Phone: (410) 516-6491. E-mail: chencke2@jhu.edu.

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to treat *Toxoplasma* as a potential substitute for the sulfa combination therapy. Because of the close resemblance of such compounds to ubiquinone, a suggested mechanism of action for atovaquone and related medications involves disruption of the mitochondrial membrane potential.[11]

Recently developed *Toxoplasma* inhibitors have shown a broad range of efficacy, toxicity, and therapeutic index (TI[a]).[12−19] The TI is a measure of tolerability of a drug expressed as a ratio of the median cytotoxic dose (TD_{50}) divided by the median inhibitory concentration (ID_{50}). The limited efficacy of current therapies due to patient drug tolerance and the relatively large quantities of drug(s) required to treat the disease necessitates the development of nontoxic, well-tolerated alternatives. Ideally these alternatives will inhibit both the tachyzoite and bradyzoite form in all intermediate hosts and will have selectivity toward the targeted lytic stage of *Toxoplasma* with minimal effect on human host cell metabolism. Additionally, therapies which target specific stages in the parasitic life cycle provide additional possibilities for synergistic combinations as well as for clarification of the drug's mechanism of action.

Artemisinin is a sesquiterpene lactone that possesses a 1,2,4-trioxane moiety. Artemisinin was first identified as the active constituent in Qinghao (*Artemisia annua*) in 1972. The artemisinin class of drugs has shown a broad range of activity against many parasites in vitro including *Plasmodium*, *Leishmania*, *Schistosoma*, *Trypanosoma*, and *Toxoplasma*.[20,21] While the 1,2,4-trioxane function has been shown to be the pharmacophore responsible for the potent activity against malaria parasites in vitro,[22,23] its role in anti-*Toxoplasma* activity is not clear. Deoxyartemisinins lacking the 1,2,4-trioxane moiety are unable to effectively block intracellular *Toxoplasma* tachyzoite replication but can moderately inhibit extracellular tachyzoite invasion of host cells.[24] Thus the illumination of the exact mechanism of action of artemisinins against in vitro *Toxoplasma* is still an area of intense interest and investigation.[25−28] Additionally, thiazole-containing compounds have been shown, in vitro, to possess antimicrobial[29] and antiparasitic properties.[30,31] We have previously shown, in vitro, that some C9−C10 dehydroartemsinin (DART) derivatives, including one possessing a thiazole moiety, inhibit multiple steps in the lytic cycle of *T. gondii*.[24] This report describes results with novel DART-thiazole, DART-oxadiazole, and DART-carboxamide derivatives.

Results and Discussion

Scheme 1 outlines the preparation of the 23 DART derivatives starting from the natural trioxane artemisinin. The specific C-10 functional groups in this study were chosen based largely on their chemical accessibility. The syntheses of the DART-1,3-thiazole derivatives were accomplished in a three-step, one-pot procedure while the DART-1,2,4-oxadiazoles and DART-carboxamides were prepared in four or five chemical steps. The DART-1,3-thiazole series were prepared by the addition to artemisinin of the lithium thiazole species, from the respective 2-bromo-1,3-thiazole that was synthesized from the corresponding aryl or alkyl ketone using published procedures.[32] The resulting thiazolyl alcohol was acetylated and then acetic acid was eliminated, in situ. The DART-1,2,4-oxadiazole series was prepared from the DART-acid[33] via the amidoxime,[34] followed by cyclization with tetrabutylammonium fluoride (TBAF).[35] The DART-carboxamide series was prepared from this same DART-acid and the requisite amine via *N*-(3-dimethylaminopropyl)-*N*′-ethylcarbodiimide (EDC)/hydroxybenzotriazole (HOBt) amide coupling.

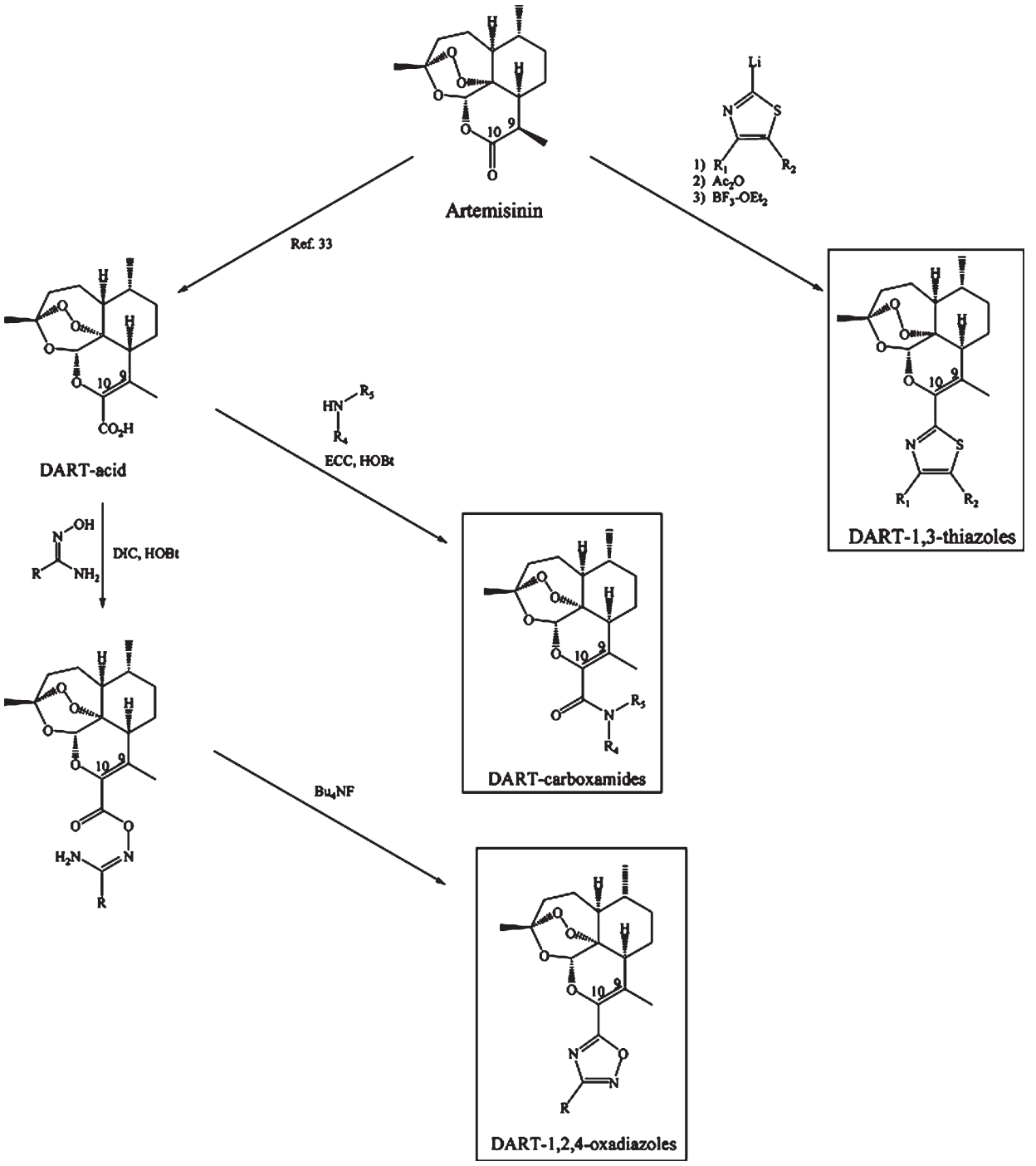
For all the prepared DART derivatives, the in vitro inhibition of growth, invasion, and replication were determined using published methods[24] (see Experimental Section). In the five-day growth inhibition assay wherein HFF (human foreskin fibroblast) cells are grown in the presence of test compounds and tachyzoites for five days (Tables 1−4), 21 of the 23 compounds displayed at least moderate inhibition (log TI $>$ 1.4) of the parasite, greater than that of trimethoprim, with 12 displaying very strong inhibition (log TI $\geq$ 2.9). These 12 compounds inhibited the parasite at a therapeutic index level similar to that of the widely used, antiparasitic trioxane drugs artemisinin and artemether (log TI = 2.9 and 3.1, respectively). Of the 23 prepared compounds, only two (**8** and **16**) were ineffective in this assay as anti-*Toxoplasma* agents owing to their cytotoxicity and thus were not investigated further in the secondary assays described below. Additionally, *N*-methyl amide **17** was not subjected to further assays because it was not more active than the corresponding *N*-ethyl amide **15**.

T. gondii establishes its lytic growth cycle in host cells through the active process of invasion. This process is initiated by attachment of the tachyzoite to the cell and ordinarily quickly progresses to penetration of that cell. The red/green invasion assay tests the ability of compounds to affect the attachment and/or penetration steps of host cell invasion. In this assay, cell-free tachyzoites are incubated with test compounds for 20 min before being added to host HFF cell monolayers. Sequential fluorescent staining of the monolayers results in extracellular/attached parasites labeled red and intracellular/penetrated parasites labeled green. (Figure 1) Thus this assay demarcates compounds that can act extracellularly, i.e., directly on the parasite, from those that require being in the intracellular environment, i.e., inside the host cell, in order to manifest antiparasitic activity. Generally, the DART-thiazoles appear to have the ability to operate extracellularly, whereas the DART-oxadiazoles and DART-carboxamides do not (Figure 1). Specifically, compounds **1**−**3** and **7**−**12** caused significant reductions in the number of penetrated (green) parasites. In this assay, an effect on attachment of parasites to host cells is defined as a decrease of numbers of both penetrated (green) and attached (red) parasites relative to same of the vehicle.[36] As shown in Figure 1, only the DART-thiazoles **1**−**3** and **7**−**12** exerted significant inhibition of attachment. It is important to note that the inhibition of attachment and the inhibition of penetration are not necessarily interrelated. It is possible to inhibit penetration while not inhibiting attachment as illustrated by cytochalasin D, a powerful inhibitor of penetration (Figure 1).

The replication assay is a relatively short duration assay that helps determine the time required for activity onset as well as the compound's ability to inhibit an established,

[a] Abbreviations: DART, dehydroartemisinin; IC_{50}, median inhibitory concentration; TD_{50}, median cytotoxic dose; TI, therapeutic index (calculated as TD_{50}/IC_{50}); TBAF, tetrabutylammonium fluoride; EDC, *N*-(3-dimethylaminopropyl)-*N*′-ethylcarbodiimide; HOBt, hydroxybenzotriazole; HFF, human foreskin fibroblast; NMR, nuclear magnetic resonance; HRMS-FAB, high resolution mass spectrometry−fast atom bombardment; PEG, polyethylene glycol; PEGMME, polyethylene glycol monomethyl ether; FT-IR, Fourier transform-infrared; TLC, thin layer chromatography; RT, room temperature; CPRG, chlorophenol red-β-D-galactopyranoside; VHL, vehicle; DAPI, 4′,6-diamidino-2-phenylindole; DMSO, dimethylsulfoxide; BAPTA-AM, 1,2-bis(2-aminophenoxy)ethane-*N*,*N*,*N*′,*N*′-tetraacetic acid tetrakis-(acetoxymethyl ester).

Scheme 1

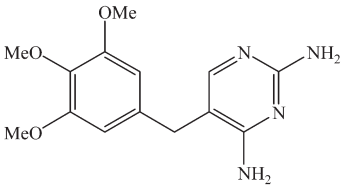


intracellular infection. All of the newly synthesized derivatives subjected to the replication assay inhibited replication to varying degrees (Figure 2). Specifically, trioxanes **13**, **15**, **18**, **21**, **23**, and **24** inhibited replication of the parasite by 50%, while trioxanes **1**−**10**, **12**, **14**, **19**, **20**, and **22** inhibited replication by 75%. Trioxanes **11** and **25** completely inhibited replication, prohibiting even one round of replication, i.e., one parasite doubling. Generally then, the DART-thiazoles appear to act quickly with all members of this thiazole series restraining replication to ≤1 parasite doubling. On the other hand, only five of the 11 DART-oxadiazole and carboxamide series (**14**, **19**, **20**, **22**, and **25**) effectively inhibited replication. By comparison, parent trioxane drug artemisinin showed no measurable activity and artemether showed only modest inhibition.

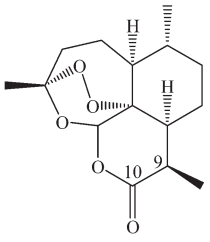
In separate experiments, compounds **2**−**6** and **14** displayed effective replication inhibition when tested over a range of concentrations (1, 5, and 10 μM), allowing no more than two parasite doublings at any concentration. Thiazoles **2**, **3**, and **4** showed the same activity at both 5 and 10 μM, with only one doubling/vacuole. However, at 1 μM, DART thiazoles **2** and **4** allowed two doublings, while thiazole **3** continued to allow just one doubling. Thiazole **6** displayed a shallow dose−response curve, never allowing more than two doublings. Further, in these dose−response experiments, both compounds **5** and **14** limited replication to 0.5 doubling at 10 μM and one doubling at both 5 and 1 μM.[37]

Several structure−activity relationships (SAR) became apparent. Monomethyl thiazoles **2** and **4** had higher therapeutic

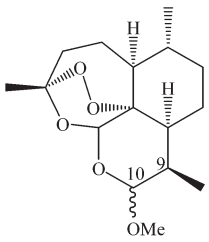
Table 1. Standards



Trimethoprim



Artemisinin

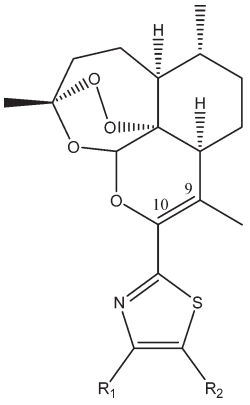


Artemether

compd	IC_{50}[a] (μM)	TD_{50}[a] (μM)	TI[a]	Log TI
trimethoprim	46	≥320[b]	12	1.1
artemisinin	0.64	≥320	879	2.9
artemether	0.31	≥320	1814	3.2

[a] IC_{50}, median inhibitory concentration; TD_{50}, median cytotoxic dose; TI, therapeutic index determined by TD_{50}/IC_{50}. [b] Cytotoxicity end point not reached; a value of $^1/_4$ $\log_{10}$ greater than the highest concentration tested was used to compute the TI.

Table 2. DART-1,3-Thiazoles

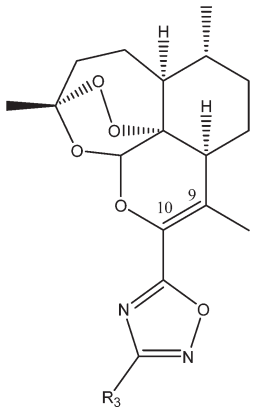


compd	R_1	R_2	IC_{50} (μM)[a]	TD_{50} (μM)[a]	TI[a]	Log TI
1[24]	H	H	1.1	≥320[b]	511	2.7
2	Me	H	0.40	≥320	1406	3.1
3	Me	Me	0.16	38	240	2.4
4	H	Me	0.37	≥320	1520	3.2
5	Ph	H	0.34	≥320	1654	3.2
6	*t*-Bu	H	0.25	≥320	2249	3.4
7	Et	H	0.40	≥320	1406	3.1
8	C_6H_{11}	H	0.59	1.8	3.0	0.5
9	*p*-CH_3Ph	H	0.35	≥320	1607	3.2
10	*p*-MeOPh	H	0.75	19	25	1.4
11	*p*-MeSPh	H	0.40	≥320	1406	3.1
12	*p*-MeS(O_2)Ph	H	0.42	≥320	1339	3.1

[a] IC_{50}, median inhibitory concentration; TD_{50}, median cytotoxic dose; TI, therapeutic index determined by TD_{50}/IC_{50}. [b] Cytotoxicity end point not reached; a value of $^1/_4$ $\log_{10}$ greater than the highest concentration tested was used to compute the TI.

indices than the corresponding dimethyl thiazole **3**. DART thiazoles bearing monosubstitution in the 4-position encompassed the rest of the thiazoles tested. DART thiazole **8**, a saturated cyclohexyl version of DART phenyl thiazole **5**, only weakly inhibited replication. *N*,*N*-Diethyl carboxamide **16** demonstrated that the amide nitrogen needs to bear at least one hydrogen atom otherwise activity is lost. *N*-Monobenzyl amides **18**−**25** comprised the remainder of the NH-amide class. 3,5-Dibromobenzyl NH-amide **25** showed potent parasite inhibition, as did the corresponding 3,5-difluorobenzyl NH-amide **24**. 4-Substitution of the benzyl ring with a fluorine atom was detrimental to inhibitory activity (see NH-amides **20** and **23**). 4-Trifluoromethylbenzyl NH-amide **22**, however, was considerably more potent than 4-methylbenzyl NH-amide **21**.

Table 3. DART-1,2,4-Oxadiazoles



compd	R_3	IC_{50}[a]	TD_{50}[a]	TI[a]	Log TI
13	Me	0.84	≥320[b]	669	2.8
14	Ph	0.44	141	320	2.5

[a] IC_{50}, median inhibitory concentration; TD_{50}, median cytotoxic dose; TI, therapeutic index determined by TD_{50}/IC_{50}. [b] Cytotoxicity end point not reached; a value of $^1/_4$ $\log_{10}$ greater than the highest concentration tested was used to compute the TI.

Finally, when DART thiazoles **5** and **6** were tested for their ability to permanently (i.e., irreversibly) inhibit tachyzoite replication, we found that, after 24 h of treatment of intracellular parasites followed by no treatment for 96 h, the parasites were effectively killed.[37,38] Thus, DART thiazoles **5** and **6** are parasiticidal; the standard trioxane drugs artemisinin and artemether are not parasiticidal.

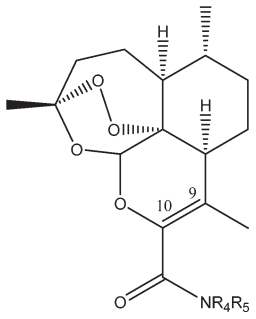
In conclusion, we report synthesis and characterization of three new classes of DART derivatives. Nearly all of the new derivatives described here are noncytotoxic, while many are also highly growth inhibitory against *T. gondii* tachyzoites, in vitro. Compounds **2**−**12** represent a new series of very potent, thiazole-containing,[19] anti-*Toxoplasma* compounds. Two of these derivatives (**5** and **6**) stand out as selective, parasiticidal compounds, a first for the artemisinin-based family of anti-*Toxoplasma* drugs. This lytic cycle stage selectivity is facilitating use of these trioxanes also as molecular probes to elucidate

the mechanism of action of these classes of DART derivatives in *T. gondii*.

Experimental Section

^{1}H NMR and ^{13}C NMR spectra were recorded on Bruker spectrometer at 400 and 100 MHz or 300 and 75 MHz, respectively, using the residual solvent peak as the internal standard. High resolution mass spectrum−fast atom bombardment (HRMS-FAB) mass spectra were obtained using a VG70SE double focusing magnetic sector mass spectrometer (VG Analytical, Manchester, UK now Micromass/Waters) equipped with a Cs^+ ion gun (28 kV @ 2 μA), an off-axis multiplier and a MSS data system (MasCom, Bremen, Germany). The resolution of the instrument was set at 10000 (100 ppm peak width). Samples were mixed with *m*-nitrobenzyl alcohol matrix deposited on the target of a direct insertion probe for introduction into the source. For accurate mass measurements, a mass scan range was employed with the matrix containing 10% polyethylene glycol (PEG) or polyethylene glycol, monomethyl ether (PEGMME) mass calibrant. Fourier transform-infrared (FT-IR) experiments were performed on a Bruker Vector 22 instrument. Optical rotation values were obtained using a 100 mm quartz cell on a JASCO P-1010 polarimeter with a 589 nm source. Thin-layer chromatography was performed with Silicycle glass-backed 20 cm ×20 cm extra-hard layer 250 μm thickness 60 Å plates with F_{254} indicator cut down to 20 mm × 67 mm for analytical purposes. The purity of all DART trioxanes was determined to be >95% by analytical thin-layer chromatography (TLC).

Table 4. DART-Carboxamides



compd	R_4	R_5	IC_{50}[a]	TD_{50}[a]	TI[a]	Log TI
15[24]	H	Et	6.8	259	38	1.6
16	Et	Et	11	23	2.1	0.3
17	H	Me	15	≥320[b]	37	1.6
18	H	Bn	0.67	≥320	839	2.9
19	H	4-methiazole	0.65	≥320	865	2.9
20	H	*p*-FBn	0.47	72	153	2.2
21	H	p-CH_3Bn	0.51	169	331	2.5
22	H	p-CF_3Bn	0.34	≥320	1654	3.2
23	H	3,4-F_2Bn	0.47	78	166	2.2
24	H	3,5-F_2Bn	0.36	≥320	1562	3.2
25	H	3,5-Br_2Bn	0.81	≥320	694	2.8

[a] IC_{50}, median inhibitory concentration; TD_{50}, median cytotoxic dose; TI, therapeutic index determined by TD_{50}/IC_{50}. [b] Cytotoxicity end point not reached; a value of $^1/_4$ $\log_{10}$ greater than the highest concentration tested was used to compute the TI.

General Synthesis of Compounds 4−12. To an oven-dried vial were added the 4-substituted 2-bromothiazole (0.62 mmol) and tetrahydrofuran (500 μL). The resulting solution was cooled to −78 °C. *n*-Butyllithium (1.6 M in hexanes, 0.62 mmol) was added dropwise, via syringe, over five min. Once the addition was complete, the reaction was stirred at −78 °C for 30 min. Then a solution of artemisinin (0.45 mmol) in tetrahydrofuran (500 μL) was added dropwise over 5 min. The resulting reaction mixture was stirred at −78 °C for 30 min and then at −65 °C for 30 min. Then acetic anhydride (3.54 mmol) was added to the reaction dropwise over 5 min, and the reaction was allowed to warm to 0 °C and to stir at 0 °C for 10 min. Boron trifluoride etherate (3.97 mmol) was added, dropwise, over 5 min, and the reaction was allowed to warm to room temperature (RT) and stir at RT for 16 h. The reaction was placed in a separatory funnel containing dichloromethane (20 mL) and saturated

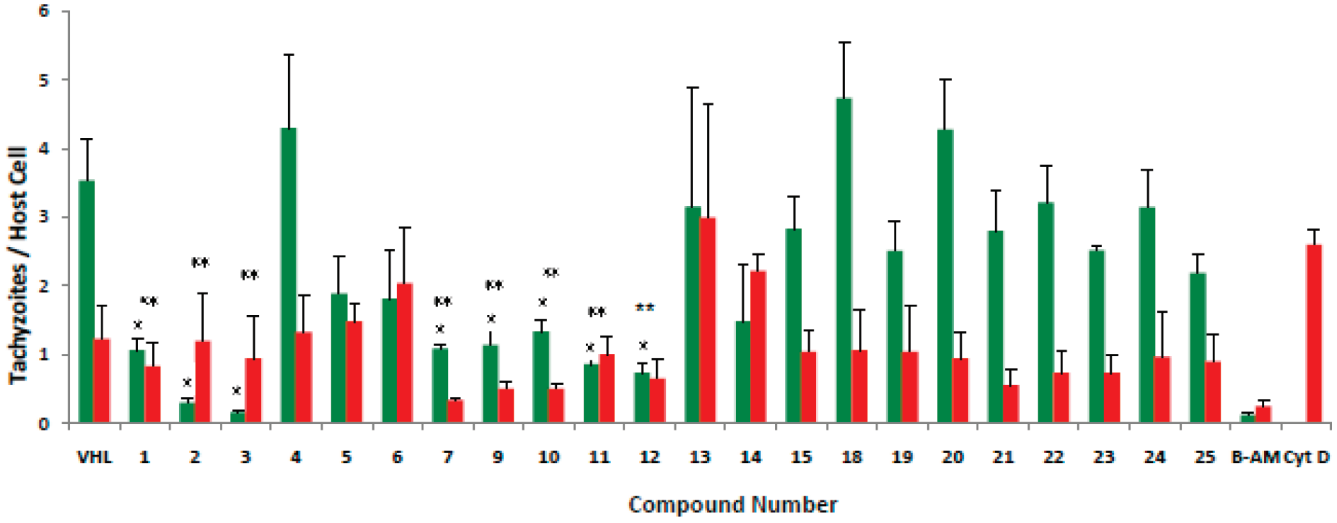


Figure 1. Quantification of invasion/attachment inhibition using red/green assay. The red/green invasion assay was performed using published methods.[24] Tachyzoites (5×10^6) were mixed with DMSO [vehicle (VHL)] or compound (final 10 μM) and allowed to sit at room temperature for 20 min before being added to HFF monolayers growing in 8-well chamber slides. After 1 h at 37 °C, 5% CO_2, the cells were rinsed and fixed. Attached/extracellular parasites were detected using Rb anti-p30 (SAG1) (AbD Serotec, UK) followed by Alexa Fluoro 594 (red) (Invitrogen, CA). After permeabilization, penetrated/intracellular parasites were stained with mAb 9e11 anti-SAG1 (Argene Inc., NY) followed by Gt antimouse Alexa Fluoro 488 (green). DAPI (Invitrogen) for staining nuclei was added to secondary antibody. BAPTA-AM (20 μM) and cytochalasin D (2 μM) are included as controls for defects in attachment and penetration, respectively.[24] Stained cells were examined by phase contrast and reflected fluorescence using an Olympus BX41 microscope. Numbers of green and red tachyzoites per host cell were enumerated by visual counting. Data are mean values ± SEM of three independent experiments, counting 10 random fields per well at 600× magnification. A single asterisk indicates tachyzoite penetration significantly lower ($P \leq 0.05$, two-tailed Students' *t*-test) than vehicle. A double asterisk indicates a significant effect ($P \leq 0.05$, one-tailed Students' *t*-test) on parasite attachment relative to vehicle.

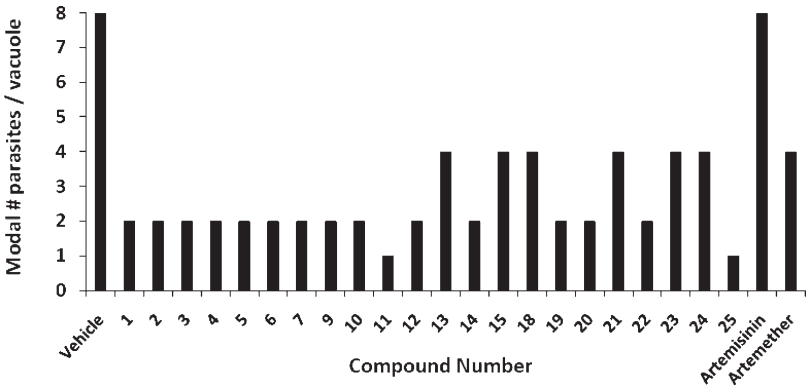


Figure 2. Quantification of replication inhibition using replication assay. Compounds were tested for replication inhibition using established procedures.[24] HFF monolayers were inoculated with tachyzoites and then incubated (37 °C, 5% CO_2) for 2 h. Compounds (final 10 μM) or DMSO (vehicle) were then added to the medium. Parasite replication proceeded for 24−26 h after which time the monolayers were fixed, permeablilized, and immunolabeled with Rb anti-p30 followed by Gt antirabbit Alexa Fluoro 594 (red). DAPI was added to secondary antibody. Cells were examined by reflected fluorescence as in Figure 1. Data were compiled from three independent experiments, each from 10 random fields per sample. The number of vacuoles containing 1, 2, 4, or 8+ parasites/vacuole were enumerated by eye. Data are expressed as the modal number of parasites per vacuole.

aqueous sodium bicarbonate solution (10 mL). The organic layer was washed with saturated aqueous sodium bicarbonate solution (2 × 10 mL) followed by saturated aqueous sodium chloride solution (10 mL). The organic layer was dried over magnesium sulfate and was concentrated, in vacuo, at 40 °C to afford crude DART-thiazole. The crude DART-thiazole was purified by flash silica gel chromatography (100 mg of silica gel/mg of crude product).

Compound 4. Crude **4** was purified by flash silica gel chromatography (100% hexanes then 10% ethyl acetate in hexanes) to afford **4** as a white solid (47.2%). TLC one spot R_f = 0.90 (40% ethyl acetate in hexanes). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.45 (s, 1H), 5.74 (s, 1H), 2.49−2.37 (m, 4H), 2.19 (s, 3H), 2.12−2.02 (m, 2H), 1.96−1.88 (m, 2H), 1.73−1.69 (m, 1H), 1.62−1.24 (m, 7H), 1.18−1.09 (m, 1H), 1.03−0.96 (m, 3H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 140.4, 138.0, 110.0, 104.5, 90.2, 78.4, 50.7, 48.0, 37.6, 36.1, 34.2, 29.0, 25.6, 24.5, 20.1, 17.1, 11.8. HRMS-FAB (m/z): $[M + H]^+$ calcd for $C_{19}H_{26}NO_4S$, 364.1577; found, 364.1581; $[\alpha]_D^{25}$ + 53.6 (c 0.640, $CHCl_3$). FT-IR (NaCl, thin film) (cm^{-1}) 2925, 2891, 1653, 1520, 1445, 1408, 1367, 1314, 1267, 1234, 1215, 1191, 1168, 1143, 1117, 1091, 1062, 1025, 1004.

Compound 5. Crude **5** was purified by flash silica gel chromatography (100% hexanes then 2% ethyl acetate in hexanes) to afford **5** as an amorphous white solid (52.2%). TLC one spot R_f = 0.72 (40% ethyl acetate in hexanes). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.96−7.94 (d, 2H, J = 7.6 Hz), 7.51 (s, 1H), 7.45−7.41 (t, 2H, J = 14.0 Hz), 7.35−7.31 (t, 1H, J = 14.8 Hz), 5.80 (s, 1H), 2.43−2.34 (m, 4H), 2.16−2.04 (m, 2H), 2.00−1.93 (m, 2H), 1.76−1.70 (m, 1H), 1.63−1.56 (m, 1H), 1.50−1.41 (m, 5H), 1.35−1.30 (m, 1H), 1.22−1.13 (m, 1H), 1.02−0.98 (m, 3H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 165.0, 155.7, 138.3, 135.2, 129.0, 128.3, 126.6, 113.5, 112.6, 111.8, 105.0, 90.9, 78.8, 51.1, 48.4, 38.0, 36.5, 34.5, 29.4, 25.8, 24.8, 20.2, 17.4. HRMS-FAB (m/z): $[M + H]^+$ calcd for $C_{24}H_{28}NO_4S$, 426.1739; found, 426.1729; $[\alpha]_D^{25}$ + 55.5 (c 38.0, $CHCl_3$). FT-IR (NaCl, thin film) (cm^{-1}) 2927, 2863, 1653, 1606, 1496, 1449, 1379, 1346, 1281, 1235, 1211, 1141, 1105, 1066, 1001.

Compound 6. Crude **6** was purified by flash silica gel chromatography (100% hexanes then 2% ethyl acetate in hexanes) to afford **6** as an amorphous white solid (54.8%) TLC one spot R_f = 0.81 (20% ethyl acetate in hexanes). ^{1}H NMR (400 MHz, $CDCl_3$): δ 6.82 (s, 1H), 5.75 (s, 1H), 2.46−2.39 (m, 1H), 2.28 (s, 3H), 2.12−2.02 (m, 2H), 1.98−1.90 (m, 2H), 1.74−1.70 (m, 1H), 1.63−1.57 (m, 1H), 1.44 (s, 4H), 1.34 (s, 10H), 1.33−1.09 (m, 1H), 0.99−0.98 (m, 3H).; ^{13}C NMR (100 MHz, $CDCl_3$): δ 166.7, 163.6, 138.2, 110.1, 109.6, 104.5, 90.2, 78.5, 50.7, 48.1, 37.6, 36.1, 34.8, 34.2, 30.1, 29.1, 25.7, 24.5, 20.2, 17.2. HRMS-FAB (m/z): $[M + H]^+$ calcd for $C_{22}H_{32}NO_4S$, 406.2052; found, 406.2048; $[\alpha]_D^{25}$ + 72.2 (c 28.5, $CHCl_3$). FT-IR (NaCl, thin film) (cm^{-1}) 2962, 2925, 2865, 1745, 1655, 1506, 1451, 1374, 1361, 1350, 1281, 1237, 1212, 1142, 1104, 1042, 1001.

Compound 7. Crude **7** was purified by flash silica gel chromatography (100% hexanes then 15% ethyl ether in petroleum ether) to afford **7** as a white foam (71.8%) TLC one spot R_f = 0.57 (20% ethyl acetate in hexanes). ^{1}H NMR (400 MHz, $CDCl_3$): δ 6.86 (s, 1H), 5.75 (s, 1H), 2.82−2.75 (q, 2H, J = 7.5 Hz), 2.42−2.32 (m, 1H), 2.21 (s, 3H), 2.12−2.02 (m, 2H), 2.00−1.88 (m, 2H), 1.73−1.67 (m, 1H), 1.63−1.44 (m, 3H), 1.40 (s, 3H), 1.32−1.27 (t, 3H, J = 7.5 Hz), 1.18−1.11 (m, 1H), 1.00−0.98 (m, 3H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 164.2, 159.6, 138.4, 112.0, 110.8, 104.8, 90.7, 78.8, 51.0, 48.2, 37.9, 36.4, 34.4, 29.4, 25.8, 25.4, 24.8, 20.2, 17.1, 13.7. HRMS-FAB (m/z): $[M + H]^+$ calcd for $C_{20}H_{28}NO_4S$, 378.1739; found, 378.1732; $[\alpha]_D^{25}$ + 61.3 (c 51.0, $CHCl_3$). FT-IR (NaCl, thin film) (cm^{-1}) 2967, 2929, 2872, 1656, 1515, 1450, 1382, 1349, 1280, 1233, 1213, 1135, 1106, 1042, 1004.

Compound 9. Crude **9** was purified by flash silica gel chromatography (100% hexanes then 2.5% ethyl acetate in hexanes) to afford **9** as an amorphous white solid (68%) TLC one spot R_f = 0.51 (20% ethyl acetate in hexanes). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.83−7.81 (d, 2H, J = 7.6 Hz), 7.39 (s, 1H), 7.23−7.21 (d, 2H, J = 7.6 Hz), 5.78 (s, 1H), 2.43−2.37 (m, 7H), 2.14−2.05 (m, 2H), 2.00−1.95 (m, 2H), 1.75−1.72 (m, 1H), 1.56−1.41 (m, 5H), 1.36−1.26 (m, 1H), 1.21−1.10 (m, 1H), 1.01 (m, 3H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 164.5, 155.4, 138.1, 137.6, 132.2, 129.3, 126.8, 126.2, 111.4, 110.9, 104.6, 90.3, 84.7, 78.4, 50.7, 48.1, 37.6, 36.1, 34.2, 29.1, 25.7, 24.5, 21.3, 20.2, 17.3. HRMS-FAB (m/z): $[M + H]^+$ calcd for $C_{25}H_{30}NO_4S$, 440.1890; found, 440.1888; $[\alpha]_D^{25}$ + 56.6 (c 0.800, $CHCl_3$). FT-IR (NaCl, thin film) (cm^{-1}) 3107, 2925, 2870, 1653, 1493, 1455, 1376, 1347, 1322, 1277, 1240, 1222, 1181, 1157, 1134, 1108, 1084, 1059, 1035, 1003.

Compound 11. Crude **11** was purified by flash silica gel chromatography (100% hexanes then 4−6% ethyl acetate in hexanes) to afford **11** as a white foam (64%). TLC one spot R_f = 0.53 (20% ethyl acetate in hexanes). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.87−7.85 (d, 2H, J = 8.34 Hz), 7.40 (s, 1 H); 7.31−7.28 (d, 2 H, J = 8.34 Hz), 5.78 (s, 1 H), 2.51 (s, 3 H), 2.46−2.39 (m, 1 H), 2.36 (s, 3 H), 2.14−2.04 (m, 2 H), 1.98−1.94 (m, 2 H), 1.75−1.71 (m, 1 H), 1.64−1.59 (m, 1 H), 1.58−1.53 (m, 1H), 1.50−1.46 (m, 3 H, including singlet at 1.46), 1.39−1.26 (m, 2H, including singlet at 1.26), 1.21−1.12 (m, 1 H), 1.02−1.00 (d, 3 H, J = 5.81 Hz). ^{13}C NMR (100 MHz, $CDCl_3$): δ 164.7, 162.4, 138.0, 126.7, 126.6, 111.7, 111.1, 104.6, 90.3, 78.4, 50.7, 48.1,

37.6, 36.1, 34.2, 33.0, 29.7, 29.1, 25.7, 25.6, 24.5, 24.5, 20.1, 17.4, 15.8. HRMS-FAB (m/z): $[M + H]^+$ calcd for $C_{25}H_{30}NO_4S_2$, 472.1616; found, 472.1610; $[a]_D^{25} + 16.2$ (c 0.32, $CHCl_3$). FT-IR (NaCl, thin film) (cm^{-1}) 2922, 2849, 1761, 1652, 1558, 1539, 1489, 1456, 1221, 1108, 1058, 1002, 919, 879, 828, 751, 665.

General Synthesis of Compound 13 and 14. A solution of DART-acid (0.15 mmol) in dichloromethane (1 mL) was added over 20 min to a solution of *N*,*N*′-diisopropylcarbodiimide (0.18 mmol), HOBt (0.18 mmol), and the substituted *N*-hydroxyalkylimidamide (0.26 mmol) in dichloromethane (3 mL) at −10 °C. The reaction was stirred for 20 min at this temperature and then overnight at room temperature. The solvent was removed under reduced pressure, and the residue was dissolved in ethyl acetate. The organic layer was subsequently washed with a saturated sodium bicarbonate solution, water, a 0.5 M potassium bisulfate solution (2 × mL) and saturated sodium chloride solution. The organics were dried over magnesium sulfate. A 1 M TBAF solution in tetrahydrofuran (218 μL, 0.21 mmol) was added dropwise to a solution of acetimidamide (62 mg, 0.15 mmol) in tetrahydrofuran (2 mL). The reaction mixture was stirred for 5 h. The mixture was poured in ethyl acetate (10 mL), washed with water and saturated sodium chloride solution, and dried over magnesium sulfate.

Compound 14. The crude product was purified by flash silica gel chromatography (11% ethyl acetate in hexanes) to afford **14** as a amorphous white solid (82% overall) TLC one spot R_f = 0.30 (20% ethyl acetate in hexanes). ^{1}H NMR (400 MHz, CD_2Cl_2): δ 8.11−8.13 (m, 2H), 7.48 (m, 3H), 5.81 (s, 1H), 2.35−2.40 (m, 1H), 2.25 (s, 3H), 1.95−2.15 (m, 4H), 1.75 (dq, J = 12.0 Hz, 4.0 Hz, 1H), 1.46−1.61 (m, 3H), 1.43 (s, 3H), 1.32 (dq, J = 12.0 Hz, 4.0 Hz, 1H), 1.15−1.25 (m, 1H), 1.01 (d, J = 8.00 Hz, 3H). ^{13}C NMR (100 MHz, CD_2Cl_2): δ 172.4, 168.7, 133.0, 131.7, 129.4 (2C), 128.0 (2C), 127.6, 120.9, 105.4, 91.0, 78.8, 51.3, 48.3, 38.1, 36.6, 35.4, 29.6, 26.0, 24.9, 20.4, 17.9. HRMS-FAB (m/z): $[M + H]^+$ calcd for $C_{23}H_{27}N_2O_9$, 411.1920; found, 411.1931; $[\alpha]_D^{20} + 32.3$ (c 0.98, CH_2Cl_2). FT-IR (KBr) (cm^{-1}) 3067, 2928, 2872, 1658, 1562, 1446, 1364, 1348, 1268, 1252, 1221, 1207, 1181, 1158, 1127, 1114, 1184, 1037, 1003, 968, 880, 830.

General Procedure for the Synthesis of Compounds 16−25. Into a flame-dried 5 mL RBF was charged DART-acid (0.10 mmol), EDC, (0.16 mmol), and HOBt, (0.12 mmol). Dichloromethane (2.5 mL) was then added, and the mixture was stirred for an hour, at which time the substituted amine (0.38 mmol) was added. The reaction was allowed to stir at room temperature for 3 h. It was then quenched with 1N hydrochloric acid, extracted with dichloromethane (3 × 5 mL), washed with saturated sodium chloride solution, dried over magnesium sulfate, and evaporated. The crude product was purified by preparative thin layer chromatography.

Compound 22. The crude product was purified by preparative thin layer chromatography (silica gel, 40% ethyl acetate in hexanes) to afford **22** as an amorphous white solid (69%). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.56 (m, 2H), 7.40 (m, 2H), 7.14 (m, 1H), 5.64 (s, 1H), 4.55 (m, 2H), 2.38 (m, 1H), 2.16 (s, 3H), 2.03 (m, 2H), 1.94 (m, 1H), 1.84 (m, 1H), 1.69 (m, 1H), 1.54−1.37 (m, 6H, including singlet at 1.37), 1.29−1.07 (m, 2H), 0.97 (d, 3H, J = 6.0 Hz). ^{13}C NMR (100 MHz, $CDCl_3$): δ 163.30, 142.80, 136.28, 127.60, 125.42, 118.21, 104.65, 90.39, 78.17, 50.43, 48.29, 42.26, 37.56, 35.97, 34.05, 28.75, 25.68, 24.28, 20.00, 16.89; $[\alpha]_D^{28} + 93$ (c 1.4, $CHCl_3$). HRMS-FAB (m/z): $[M + Na]^+$ calcd for $C_{24}H_{28}F_3NO_5Na$, 490.1812; found, 490.1814. IR (NaCl, thin film) 3441, 2930, 2873, 1717, 1653, 1619, 1558, 1540, 1508, 1456, 1418, 1377, 1326, 1219, 1161, 1128, 1067, 1036, 1018, 1001, 964, 924, 881, 831 cm^{-1}.

Compound 24. The crude product was purified by preparative thin layer chromatography (silica gel, 40% ethyl acetate in hexanes) to afford **24** as an amorphous, white solid (61%). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.12 (m, 1H), 6.81 (m, 2H), 6.65 (m, 1H), 5.65 (s, 1H), 4.47 (m, 2H), 2.39 (m, 1H), 2.15 (s, 3H), 2.02 (m, 2H), 1.94 (m, 1H), 1.82 (m, 1H), 1.70 (m, 1H), 1.57−1.38 (m, 6H, including singlet at 1.38), 1.27−1.06 (m, 2H), 0.98 (d, 3H, J = 5.2 Hz). ^{13}C NMR (100 MHz, $CDCl_3$): δ 163.27, 142.89, 136.24, 118.39, 110.07, 109.88, 102.47, 90.43, 78.16, 53.35, 50.45, 48.32, 41.96, 37.57, 36.00, 34.07, 28.73, 25.66, 24.30, 20.01, 16.88; $[\alpha]_D^{24} + 99$ (c 0.25, $CHCl_3$). HRMS-FAB (m/z): $[M + H]^+$ calcd for $C_{23}H_{28}F_2NO_5$, 436.1936; found, 436.1921. IR (NaCl, thin film) 3442, 2926, 2876, 1653, 1625, 1597, 1558, 1508, 1458, 1376, 1316, 1279, 1218, 1118, 1100, 1036, 1004, 924, 880, 832 cm^{-1}.

Five-Day Growth Inhibition Assay. The five-day growth inhibition assay was used to screen a test compound's ability to inhibit parasite growth, both intracellularly and extracellularly. Tests were performed using human foreskin fibroblast (HFF; ATCC, VA) host cells plated in 96-well plates and tachyzoites of *T. gondii* strain 2F (ATCC) which constitutively express β-galactosidase (β-gal). Test and control compounds were added to the first column of cells (final 320 μM) and then serially diluted across the plate by dilutions of 0.5 log10, leaving the final column drug-free (0 μM) (parasite control). Following this, tachyzoites were added to the top six rows of wells, thus leaving the bottom two rows uninfected for cytotoxicity determinations. After 4 days of incubation at 37 °C/5% CO_2, the substrate for β-gal, chlorophenol red-β-D-galactopyranoside (CPRG), was added to the *Toxoplasma* wells and the plates incubated for one additional day. The cell viability reagent, CellTiter 96Aqueous One solution reagent (Promega Corp., WI) was then added to the bottom two rows of the plate. After three hours of incubation, color reactions in the wells were read in a Vmax microplate reader (Molecular Devices, CA). The amount of absorbance (570−650 nm) in wells containing drug, tachyzoites, and CPRG was compared to that in parasite control wells. The amount of absorbance in these wells is directly proportional to the amount of β-gal activity and thus to the amount of viable tachyzoites. Thus, a decrease in the amount of absorbance indicates an inhibition of enzyme activity and, by extension, parasite growth. In the cytotoxicity wells, the bioreduction of the cell viability reagent by viable cells into a soluble, colored formazan product was captured by reading the plates at 490−650 nm. The median inhibitory concentration (IC_{50}) and median cytotoxic dose (TD_{50}) were calculated using CalcuSyn software (Biosoft, Cambridge, U.K.). For each compound, a therapeutic index (TI) was calculated with the formula TI = TD_{50}/IC_{50}. Data shown (Tables 1−4) are compiled from results from at least three independent experiments.

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Supporting Information Available: Spectral data for compounds **1**−**3**, **8**, **10**, **12**, **13**, **15**−**21**, **23**, and **25**. This material is available free of charge via the Internet at http://pubs.acs.org.

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